

ARMENISE-HARVARD SYMPOSIUM 2000

ARMENISE FOURTH SYMPOSIUM

*4th Biennial Symposium
June, 2000, Bretton Woods, New Hampshire*

About the Symposium

The Fourth Annual Symposium of the Giovanni Armenise-Harvard Foundation was held in late June in an emerald valley at the foot of Mt. Washington, the highest peak in the Northeastern United States. In this spectacular setting the Foundation held its first scientific meeting of the new millennium, which was the largest so far with more than 130 Italian and American scientists in attendance.

In his opening remarks, Dr. Stephen C. Harrison of Harvard Medical School reminded participants of the historic past of the Mt. Washington Hotel. It was here that representatives from 44 nations convened, in July 1944, for the landmark Bretton Woods Conference. With World War II drawing to a close, international financial and policy experts gathered to plan for post-war economic stability. They created the International Monetary Fund as a means for stabilizing currency and exchange rates, and the World Bank as a lender to help member countries rebuild and develop after peace was restored.

"It is therefore a totally appropriate site for a conference sponsored by a foundation set up by the generosity of a banker," said Dr. Harrison, paying tribute to Count Giovanni Auletta Armenise, who along with his late wife laid the groundwork for the Foundation in 1994. A few days before this symposium began, a building at Harvard Medical School was named for Count Armenise as a permanent reminder of his contributions to international scientific advancement.

For the past four years, the Foundation has been the catalyst for new collaborations among Harvard Medical School's basic science departments and between HMS and scientific centers in Italy. Much as financial programs born at the Bretton Woods Conference benefited millions of people during the last half of the 20th Century, international scientific cooperation now promises to make life better for future generations. Since its inception, the Foundation's philosophy has been that basic science discoveries will have far-reaching impact on fields as seemingly disparate as medicine and

agriculture. During this year's symposium, a series of presentations on the common defense strategies of plants and animals demonstrated that this promise is being fulfilled.

Findings from Armenise-supported research are shared not only during the annual symposium, but also at other conferences focusing on basic biology topics. The Foundation also underwrites a broad range of joint ventures, including exchanges of technology and personnel. Five leading Italian institutions conduct Armenise-sponsored research: the European Institute of Oncology in Milano, the University of Padova, the Institute for Cancer Research and Treatment at the University of Torino School of Medicine, the Dipartimento di Ricerca Biologica e Tecnologica (DIBIT) at Scientific Institute San Raffaele in Milano, and Universita' Di Roma La Sapienza. At HMS, the four Armenise centers are structural biology, neurobiology, cell signal transduction, and human cancer viruses. All were represented on this year's scientific program.

The Fourth Annual Symposium featured 20 formal lectures and 32 posters grouped into five sessions:

- Signal Transduction, Oncogenes, Development
- Cellular Differentiation
- Activities of Nerve and Muscle
- Pathogens and Defense
- Cell Cycle, Senescence, Programmed Cell Death

The organization of this report mirrors the symposium program. Each section begins with introductory remarks on the general topic, followed by summaries of individual presentations

Session 1: Signal Transduction, Oncogenes, Development

Overview

These presentations illustrate how signal transduction has snowballed over the past decade from a relatively narrow field into a broad discipline that touches nearly every facet of cell biology and medicine. In earlier times, a typical signal transduction experiment examined the effects of growth factors on cells in culture, Dr. Stephen C. Harrison said when he introduced this session. "Then, groups of people suddenly realized they were working on the same thing." Cancer cell biologists who originally explored the relay of signals within cells were joined by developmental biologists who studied the regulation of form and pattern and the differentiation of cells into structures such as skin, hair, or heart. More recently, clinical investigators jumped on the signal transduction bandwagon as they realized that this field might unlock an array of human diseases.

The four papers in this session represented a wide spectrum: a classic look at oncogenic conversion, followed by two studies of cytoskeleton proteins, and finally an examination of signaling's role in the architecture of the heart. Each offered a detailed analysis of the function and structure of some of the players in some of the pathways that help determine larger phenomena. A common theme of these presentations, in Dr. Harrison's view, was that "signal transduction is taking over studies of the cytoskeleton." Without the microtubules, actin filaments, and other proteins of the cytoskeleton, eukaryotic cells would be shapeless and immobile. Cells require a whole library of elaborate computer codes to go about their business, and Dr. Harrison described signal transduction events as "microscopic subroutines within those codes."

Presentations

Molecular mechanisms underlying oncogenic conversion of scatter factor receptor

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Hepatocyte growth factor (HGF) is one of several scatter factors that play a pivotal role in normal embryonic development. These scatter factors stimulate branched morphogenesis, a process essential for formation of neural, epithelial, and some mesodermal-derived tissues, including muscles and bones. HGF's receptor is Met, encoded by a member of the MET/RON/SEA oncogene family. These receptors are connected to Ras, a signaling pathway that is an established contributor to malignancy and metastasis.

Dr. Michieli and his colleagues have been studying scatter factors and their receptors for many years, and recently they have focused on mutations in the tyrosine kinase domain of Met. Such mutations have been identified in papillary renal carcinoma, a human kidney tumor, and the researchers wanted to pin down the mutation's contribution to disease. To do this, they analyzed

the biochemical and biological properties of numerous Met mutants, and observed whether such mutations were sufficient to turn ordinary mouse fibroblasts into cancerous cells. Most of the mutants stepped up catalytic activity in the cells, and those with the greatest transforming potential had the highest kinase activity and the strongest link to signal transducers. The best transformers hyperactivated the Ras signaling pathway, while the less aggressive ones protected against apoptosis.

But there was a catch. In epithelial cells, which don't make HGF on their own, even the mutations that raised catalytic activity to the highest levels could not turn cells malignant unless recombinant HGF was added to the mix. In mouse fibroblasts, which produce HGF, numerous mutants could turn cells malignant. Transformation was easily blocked, however, by adding HGF antagonists or by using site-directed mutagenesis to keep the Met receptor from binding HGF. These data suggest that although Met mutations may have the capacity to cause cancerous changes in cells, this won't happen in the absence of HGF. It is as though they know how to dance, but will only perform when they hear music. *In vivo*, the kidney and liver have abundant HGF, which makes it highly likely that the Met mutations found in human kidney tumors are indeed pathogenic.

From Ras to Rac: not just a matter of guanine nucleotide exchanges (GEFs)

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Growth factors set off a series of chain reactions in cells, and while one regulates cell cycle a second may be molding the shape of the cell itself. Dr. Scita's laboratory has examined the cross-talk between the Ras pathway and Rac, a small guanine nucleotide (GTP)-binding protein that is a crucial organizer of the actin cytoskeleton. Rac has been identified as a key downstream target in Ras signaling, and the researchers performed a series of experiments aimed at identifying intermediaries that carry signals between the two. One player is a substrate of receptor tyrosine kinases called Eps8, which binds to a protein designated E3b1/Abi-1. Dr. Scita's team recently showed that Eps8 and E3b1/Abi-1 participate in the transduction of signals from Ras to Rac by regulating Rac-specific guanine nucleotide exchange (GEF) activities.

The plot thickened when the researchers realized that *in vivo*, Eps8 and E3b1 form a tricomplex with a GEF protein called Sos-1. When all three work together, they enable Rac to organize actin filaments into "ruffles" of the cell membrane. But if either Sos-1 or E3b1 is blocked, the ruffles don't form. Further experiments indicated that although Sos-1 acts on Rac when it is part of this tricomplex, it functions quite differently if hooked to a different partner. When the Sos-1 protein is coupled with a Grb2, an adaptor protein, it is recruited to the plasma membrane where it activates Ras by catalyzing the exchange of guanosine diphosphate for guanosine triphosphate.

Although Sos-1 is a versatile player that can function either upstream or downstream of Ras, it cannot play both roles at once. *In vitro*, it is clear that Grb2 and E3b1 compete for binding to Sos-1. *In vivo*, E3b1 overexpression kept Grb2 from associating with Sos-1 and favored the formation of the Eps8-E3b1-Sos-1 tricomplex. This complex has Rac-specific GEF. Additional experiments provided further evidence that Sos-1's specificity as a GEF depends entirely on how it is complexed: a receptor-Grb2-Sos-1 complex results in Ras activation, whereas an Eps8-E3b1-Sos-1 complex regulates Rac activation.

Recognition of a proline motif in beta-dystroglycan by an ""embedded"" WW domain in human dystrophin

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Several years ago, Dr. Eck's laboratory was the first to determine the crystal structure of the protein encoded by Src, the first human oncogene to be discovered and the flagship of the tyrosine kinase class of cell-surface receptors. Src is like a switch, made of several different components, that can be flipped by a variety of stimuli. One part of the switch is the Src homology-3 (SH3) domain, a structure that Dr. Eck's team recently found not in cancer, where one might expect to see it, but in a hereditary disease of muscle.

Dystrophin, the protein that is defective in Duchenne and Becker muscular dystrophies, serves as a scaffold for signaling molecules and forms a structural link between the actin cytoskeleton and the extracellular matrix. Dystrophin is linked to the cell membrane through a protein called beta-dystroglycan. The C-terminal region of dystrophin binds the cytoplasmic tail of beta-dystroglycan, in part through the interaction of a WW domain on dystrophin with a proline motif (PPxY) in the tail of beta-dystroglycan. This WW domain is homologous with SH3; in this setting, it is stabilized by an adjacent helical region that contains EF hand-like domains. The crystal structure of the dystrophin and beta-dystroglycan complex shows that beta-dystroglycan peptide binds a composite

surface formed by the WW domain and one EF-hand. Embedded in a larger binding molecule, the WW-domain recognizes the PPxY motif much as SH3 would do.

In a separate series of experiments, Dr. Eck's team found another Src-like structure, called SH2, in a negative regulator of signal transduction called CBL. This protein down-regulates tyrosine kinase receptors by marking them for proteolysis. The N-terminal of CBL contains an SH2 domain, again combined with an EF-hand structure, that grabs phosphorylated tyrosine kinases.

Like Legos, standard modules crop up in different settings and their functions are at least partly determined by context. These observations show how efficient nature is at reusing the same hardware for different purposes, Dr. Eck said.

How hearts are made: The genetics behind the induction and patterning of the heart field

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One of medicine's holy grails is to be able to repair damaged heart muscle. Dr. Mercola's research is predicated on the idea that if researchers knew exactly how cardiomyocytes develop in the embryo, it might eventually be possible to recreate myocardial tissue for therapeutic use.

In order to actually form a heart, prospective heart tissue must receive signals from two adjacent tissues: endoderm that will form the floor of the pharynx and dorsal midline mesoderm that will form the notochord and head mesoderm. Once induced, the heart field is then subdivided into distinct myocardial and non-myocardial compartments, in part by interactions with neurogenic tissue. Recent experiments in his laboratory have focused on two systems that mediate these processes; one involves a growth factor and the other a receptor-ligand pair.

Wnt is a diffusible growth factor, and Dr. Mercola's team has found that the dorsal midline mesoderm secretes several Wnt antagonists, such as Dkk1 and Frzb, that induce genes needed to begin turning non-cardiogenic mesoderm into myocardium. Although additional signals from the endoderm are required for progression to a heart tube, he believes that the location and extent of cardiogenic mesoderm in the embryo depends on the distribution of these endogenous Wnt antagonists. Tissue where Wnt is unopposed will not become part of the heart.

Genes encoding the transmembrane receptor Notch1 and its ligand Serrate1 are expressed in a pattern that strongly suggests they subdivide the heart field into myocardium and non-muscular components such as valves. When the Notch pathway was activated through the downstream transcription factor Su(H), myocardial gene expression was inhibited and non-myocardial genetic markers increased. When Notch and Su(H) function was blocked, mesoderm differentiated into

cardiomyocytes. Moreover, lineage analysis showed that cells where Notch signaling was activated did not contribute to myocardial tissue. Clearly, cells will only choose to become myocytes in the absence of Notch signaling.

Session 2: Cellular Differentiation

Overview

One of biology's greatest wonders is that a fertilized egg gives rise to an embryo made up of myriad cell types that are not only chemically different, but also arranged in a specific, three-dimensional pattern. Cells that inherited exactly the same genetic material from the egg diverge into brain and bone, hair and heart. Such chemical and architectural variety is possible because genes are switched on or off, and are expressed differently in diverse tissues.

This session, like the opening one, also concerned development, Dr. Tullio Pozzan said in his introduction. The first presentation focused on a novel method for identifying genes that are activated in normal and malignant growth of epithelial cells. The other three explored various facets of neuronal tissue development.

Presentations

The transcriptional response of epithelial cells to scatter factors

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In normal development, scatter factors stimulate epithelial cells to execute a complex program that culminates with polarization and the formation of tubules; in invasive tumor growth, this normal process is subverted and cells proliferate and invade in abnormal ways. One way to understand the differences between normal and malignant growth is to compare which genes are switched on in each. In order to do this, Dr. Medico's team created a ""gene trap"" - a novel fusion protein that can be used to screen a cell's entire genome for activated genes, whether or not their sequence and function are known.

They built a promoterless retroviral vector carrying a reverse-oriented splice acceptor (or ROSA) gene and the sequence for a green fluorescent nitro-reductase (GFNR) fusion protein that serves as a marker. When this gene trap encounters an active promoter, the trap construct will integrate itself downstream and the marker will be expressed. Cells that have taken up the trap can then be identified using FACS analysis. Conversely, the nitro-reductase moiety allows pharmacological selection against constitutive GFNR expression. A mouse liver cell line was stimulated with hepatocyte growth factor (HGF), a scatter factor, and screened with some traps set to select for HGF-induced genes, and others designed to pick out genes suppressed by exposure to HGF. Some 60 different traps were used to categorize genes and pick out the most promising HGF targets. Several responsive clones were isolated, and regulated expression of the trapped gene was confirmed at the RNA level. When Dr. Medico and his colleagues sequenced the regions around trap sites, they found genes that had never before been linked to scatter factor biology. The goal of future studies will be to shed light on transcriptional response in normal and cancerous cells.

The establishment of neuronal identities in the developing nerve cord

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The long-range goal of Dr. Thor's work is to understand the molecular genetic mechanisms that control establishment of motor neuron identities. His laboratory uses Drosophila as its primary model, and although the fly has a relatively simple nervous system it still features about

100 distinct types of cells that can be classified as neurons, glia, or interneurons. Recent experiments have focused on LIM homeodomain proteins, a family of transcription factors that are expressed in discrete subsets of developing neurons throughout the animal kingdom.

Dr. Thor's experiments indicate that three LIM-HD genes, *islet (isl)*, *lim3*, and *apterous* act in a combinatorial code to specify motor neuron subtype identity. By attaching markers to mutant versions of the genes, he has found that they control two basic hallmarks of neuronal identity - they guide axons toward target cells and specify which neurotransmitters are turned on. Additional genes are probably required to establish the ultimate, unique identity of neurons, and current research focuses on identifying them.

Because LIM-HD programs appear to be highly conserved, Dr. Thor hopes that his findings will ultimately help medical scientists understand vertebrate motor neuron generation and differentiation. With this knowledge in hand, it may someday be possible to replace cells that are lost in spinal cord injuries or neurodegenerative disorders.

Ebf genes in vertebrate neural development

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A classic family of helix-loop-helix (HLH) transcription factors are the myogenic proteins, which are well known for their role in the differentiation of muscle cells. Dr. Consalez' lab has a long-time interest in a different subclass of transcription factors with the HLH DNA-binding motif, called the Ebfs. This gene family was originally implicated in B-cell maturation and olfactory function. Several years ago, his group identified two family members in the mouse (*Ebf2*, *Ebf3*); more recently they found two more in *Xenopus laevis* (*Xebf2*, *Xebf3*). Other investigators have cloned Ebf family members from the nematode *C. elegans*.

Just as myogenic proteins can trigger events that turn epidermal cells into myoblasts, Dr. Gonzalez' team has demonstrated that overexpression of *Ebf* genes in *Xenopus laevis* embryos can transform presumptive epidermis into neurons. Ebf expression in frogs begins very early in embryonic development and continues through the tadpole stage, with different genes acting at different times. *Xebf2* operates at early stages of neuronal differentiation, upstream of *NeuroD*, whereas *Xebf3* is a target of *NeuroD* and plays a role in terminal neuronal differentiation. In the mouse, three *Ebf* genes have been shown to advance neuronal differentiation after primary neurogenesis is underway. A tantalizing feature of Ebf proteins is that they have intrinsically different functions, and act at different stages of development, despite having very similar molecular structures.

Signal transduction pathways that regulate neuronal survival in the developing mammalian central nervous system

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For nearly 100 years, scientists have viewed the development of the central nervous system as a life and death matter. Cells are initially produced in huge excess, then whittled away by cell death as development proceeds. Which cells live or die is regulated by extracellular growth factors, such as neurotrophins, and Dr. Bonni's laboratory focuses on exactly how these life-or-death decisions are carried out. Using cerebellar granule neurons obtained from rat pups and grown in culture, he has been able to pinpoint the pro-life activities of a polypeptide growth factor called brain-derived neurotrophic factor (BDNF).

BDNF appears to promote cell survival in several ways. It activates the Ras-MAPK and PI-3 NKT cascades, which team up to modify BAD, one of a class of proteins that are known to act as gatekeepers of the cell-death machinery. In its native form, BAD promotes cell death by binding to and suppressing pro-survival members of the Bcl-2 family. BAD can no longer kill developing cells, however, if signaling proteins in the Ras-MAPK and PI-3 NKT pathways phosphorylate it at two specific sites.

In addition to this transcription-independent mechanism, Dr. Bonni found that BDNF also promotes cell survival through transcription-dependent means. The MAPK-Rsk path acts on CREB, a transcription factor known to promote cell survival. He hypothesized that while transcription-

independent mechanisms might allow cerebellar granule neurons to survive shortly after they are generated on the outer surface of the developing brain, transcription-dependent mechanisms might act later on, as these neurons differentiate and mature in the brain's interior

Session 3: Activities of Nerve and Muscle

Overview

Neuroscience is one of the broadest contemporary scientific disciplines, both in terms of its methods and what they are used to study. It encompasses everything from basic biophysics to clinical surveys aimed at linking human diseases with genetic abnormalities. Although three of the four reports in this session use molecular tools to study nerves and muscles, they nevertheless illustrate some of the field's diversity. In his introductory remarks, Dr. Elio Raviola of HMS noted that most scientific lectures about signal transduction and other intracellular events are accompanied by slides showing circles, squares, connecting lines, and directional arrows. But none of these can be seen under the microscope, he observed to appreciative laughter. In fact, little is really known about *where* many molecular events actually take place within living cells. The lead paper in this session described a technique that can be used to track specific chemical changes in real time.

The next two presentations took a molecular look at the interface between nerves and muscles. The first considered calcium's role in the flow of information from the pre-synaptic to the post-synaptic neuron, and how this influences the activity of the synaptic cell. The second examined signal

transduction pathways that link electrical signals at the muscle cell surface to transcriptional commands in its nucleus, and along the way uncovers a new role for Ras.

The narrative sweep of the final paper was unusually broad. Here the researchers had to function as social scientists in the field, making contact with members of a sprawling Italian family, before they could use the tools of molecular genetics in the laboratory. The reward was the discovery of a new mutation responsible for an unusual form of epilepsy.

Presentations

Imaging signal transduction in living cells

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Although tyrosine kinase receptor cascades are important, they are only one part of the signal transduction story. Other important players include small molecules known as intracellular mediators or second messengers, especially Ca^{2+} and cAMP. These widely used messengers receive signals from surface receptors, and transmit signals to cellular proteins. Dr. Pozzan's lab has developed novel techniques for pinpointing second messenger activity and monitoring their dynamic interactions. His team used several fluorescent proteins, produced by the jelly fish *Aequorea victoria*, to engineer specialized sensors that track the activities of Ca^{2+} or cAMP in living cells. Some of their findings challenge the conventional wisdom.

For example, the mitochondria have traditionally been seen as the main organelle involved in Ca^{2+} handling. A sensor made with the Ca^{2+} sensitive photoprotein aequorin, which lights up and "freezes" Ca^{2+} activity in stimulated cells, made it possible to localize calcium activity in subpopulations of organelles. Studies with this luminescent probe revealed that in fact a subpopulation of mitochondria, which huddle around the endoplasmic reticulum and do not stray, carry out most of the calcium exchange.

The researchers invented a second sensor that could be used to localize cAMP signaling. This probe tags the regulatory (RII) and catalytic (Cat) subunits of protein kinase A (PKA) with either of two types of green fluorescent protein. The two GFPs were selected for their ability to generate fluorescence resonance energy transfer, or FRET. PKA is the main effector of cAMP in eukaryotic cells. When cAMP is low inside the cell, the RII and Cat subunits of PKA are in close proximity and the donor GFP can transfer energy to the nearby acceptor GFP. When cAMP levels increase, cAMP binds to the RII subunit and the active Cat subunit is released. FRET disappears as soon as the two are separated. By measuring the ratio of blue to green emissions, Dr. Pozzan's team can localize and measure cAMP fluctuations in response to selective stimulation of plasma membrane receptors.

Calcium control of transmitter release during realistic activity patterns

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During the normal operation of the brain, neurons often fire in intense, high frequency bursts, separated by long silent intervals. Calcium channels open along the axon as signals travel toward the bouton where neurotransmitter is squirted into the synapse. Although synapses are known to undergo profound strength changes in response to activity, and although changing levels of calcium ions are thought to regulate the strength of the neurotransmitter message, this calcium activity has been quite difficult to observe.

Several years ago, Dr. Regehr and his colleagues developed a method for using calcium-binding fluorescent dyes to study how calcium ions govern neurotransmitter release in different types of neurons. Recently, the researchers have focused on ""climbing fiber"" synapses that drive the Purkinje cells in rat cerebellum. Some of their findings are surprising: they expected a cell that released a huge burst of neurotransmitter to recover more slowly than a cell that delivered a smaller amount. When presynaptic cells were rapidly and repeatedly stimulated, however, they recovered much more quickly than the researchers predicted. In general, the higher the calcium level, the faster they recuperated and fired again. When the researchers experimentally manipulated calcium levels in presynaptic cells, they found that both release and recovery could be altered. Although there is still much to learn about the dynamic regulation of synaptic strength, Dr.

Regehr's interpretation is that the synapses have a complex system for filtering inputs and controlling synaptic outputs during complex activity patterns.

Nerve activity-dependent regulation of the muscle phenotype: a new role for Ras

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The type of nervous stimulation that a muscle receives is an established factor in both its growth and the type of fiber it comprises. Less clear are the signal transduction pathways that link depolarization at the muscle cell's surface with transcriptional changes in its nucleus. In order to identify these pathways, Dr. Schiaffino's lab uses the rat soleus muscle, in the animal's lower limb, as an in vivo muscle regeneration model. As a result of these studies, they have found a new role for the familiar Ras signaling pathway.

Local changes at the neuromuscular junction and broader changes in muscle phenotype occur when a muscle is deprived of nerve stimulation. If an injured rat soleus muscle is not reinnervated with several days, genes that produce fast-fiber myosin will quickly predominate. New connections will

appear, but normal activity will not be restored. If natural healing takes place and the muscle is reinnervated, large quantities of slow-fiber myocin will be produced and function will return. In the laboratory, this effect can be reproduced by electrostimulation using a continuous, low-frequency pattern.

Knowing that electrical stimulation could restore normal myocin production, Dr. Schiaffino and his colleagues manipulated the Ras signal transduction pathway. They transfected regenerating muscles with either constitutively active Ras or a negative Ras mutant. An unexpected finding was that the active Ras mutant stepped up production of slow-fiber myocin and down-regulated fast myocin - even in cells that were denervated. The dominant negative Ras, in contrast, interfered with regeneration even in electrically stimulated muscles. Additional experiments showed that selective activation of different pathways downstream of Ras had differing effects on muscle growth and fiber type. RasV12S35, which activates the MAPK (ERK) pathway, was able to induce slow myocin but not muscle growth; RasV12C40, which activates the PI3K pathway, affected muscle growth but not myosin gene expression. In addition to the traditional association of Ras and inhibition of myoblast fusion and muscle cell differentiation in culture, this study identifies a new role for this pathway in the differentiation of muscle phenotype by nerve activity.

Idiopathic epilepsy: analysis of a positional candidate gene

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Epilepsy is a label applied to a broad spectrum of seizure disorders, ranging from mild and occasional to frequent and severe, that affects as many as 1 in 200 people in the general population. Partial or focal forms account for about 60% of epilepsy cases, and there is evidence that a few percent of these are due to single gene abnormalities. Two gene loci have been linked with a Mendelian form of partial epilepsy, called autosomal dominant nocturnal frontal lobe epilepsy, or ADNFLE. The typical clinical presentation involves seizures during light sleep, which are frequently misdiagnosed as nightmares. Among affected members of the same pedigree, some rarely experience episodes and others are troubled by frequent, even nightly seizures. Symptoms of ADNFLE, which typically begin in childhood and do not worsen over time, usually respond well to treatment with carbamazepine or other anti-seizure medications.

Dr. Casari's team learned of a large Italian family that included many members who, beginning around age 9 to 12, experienced seizures during sleep. Most had a prodrome that involved auras, shivering, or tingling sensations. The researchers used genome-wide linkage mapping to track a third locus for ADNFLE, which they call ENFL3, to a large region of chromosome 1. This region turned out to include CHRN2, which encodes the beta-2 subunit of the neuronal nicotinic receptor (nAChR). The researchers found that family members with ADNFLE had a mis-sense mutation in this gene that resulted in this gated channel remaining open for an abnormally long time. This finding demonstrates that ADNFLE is more genetically heterogeneous than previously thought, and indicates that the cholinergic system plays a pathogenic role in this form of partial epilepsy.

Session 4: Pathogens and Defense

Overview

Since its inception, the Armenise-Harvard Foundation has sponsored basic research on plants as well as animals, believing that advances in agriculture as well as medicine hold tremendous benefits for future generations. Thus, each year's symposium has featured lectures on pathogenesis and defense mechanisms in the plant world. This year, the synergy between plant and animal science was more apparent than ever before. Mammalian biologist Tomas Kirchhausen, who chaired the session, admitted that he knew little about plants before he began preparing for the symposium. And the Harvard researcher was astonished to learn that plant and animal defense systems not only resemble one another, but sometimes deploy the same genetic and molecular elements when battling their enemies.

The first presentation in this session, for example, concerned a plant pathogen that injects harmful proteins into its leafy victims with the same syringe-like structure that salmonella uses to invade the lining of the human gut. In the second talk, the emphasis shifted to experimental methods that can be applied to either kingdom. Here it became clear that X-ray crystallography, which has provided remarkable insights into human dramas such as the binding of HIV to T cells, can also illuminate pathogen-host interactions in plants. The third paper focused on a class of proteins that act as both sentinels and warriors, recognizing certain invaders and doing battle with them. Pathogen strategies for evading host defenses were the focus of the closing talk, which made the transition from plants to animals by concentrating on how peptide antigens are presented on cell surfaces.

Presentations

Structural studies on a fungal pathogenicity factor

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The first task of any plant-attacking microbe is to batter its way through the ramparts of the cell wall. A high resolution structural analysis of a fungal battering ram, done by Drs. Mattei and Federici, reveals that the best form of this weapon is also the most recognizable to plant defenses. Fusarium moniliforme, like other fungi, uses endopolygalacturonases (PGs) to break cell-wall proteins into pieces and dissolve them. These investigators used X-ray crystallography to determine the structure of PG from F. moniliforme at 1.73 Å resolution. Like other pectinolytic enzymes, this PG resembles a squared-off coil of spring, with 10 coils each made up of three or four parallel beta-helical strands, in a coil-coiled helical organization.

The researchers prepared an assortment of site-directed mutants of PG and used these to sort out which PG residues are involved in catalysis and which interact with defensive polygalacturonase inhibiting protein (PGIP). It appeared that three aspartic acids and one histidine may be involved in catalysis, but play no role in recognition by PGIP. The researchers found that amino acid substitutions at two residues (Lys269 and His188) interfered dramatically with the binding of PG and its inhibitor. These and other results led the investigators to hypothesize that inhibition

mechanisms involve both competition between substrate and inhibitor and the covering of an active site cleft.

Delivery of bacterial effector proteins to plant cells specifying plant disease resistance

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Xanthomonas campestris pv. vesicatoria (Xcv) causes a bacterial spot disease that can devastate susceptible pepper and tomato crops. Dr. Staskawicz' laboratory studies the match between bacterial avirulence genes and the host resistance genes that recognize them, focusing mainly on the Xcv gene avrBs2 and on Bs2, the corresponding plant resistance gene.

Their data suggest that X. campestris delivers the AvrBs2 avirulence protein via a syringe-like structure called the Hrp Type III secretion system. This mechanism is familiar to eukaryotic biologists because salmonella, shigella, and other animal pathogens use it to inject proteins into host cells. It's remarkable that plant and animal pathogens have both come up with

this machinery, Dr. Staskawicz observed, and many researchers are now analyzing its evolution and genetic underpinnings.

In the meantime, Dr. Staskawicz' findings about effector proteins made by avirulence genes and the receptors that recognize them are being put to use in the field. Years ago, agriculturists found a *Bs2* resistance gene in wild peppers and successfully bred it into commercial pepper strains. As a result, many farms grow Xcv-resistant peppers. There is no indigenous *Bs2* gene in tomatoes, however, so Dr. Staskawicz and his collaborators have used the pepper resistance gene to create transgenic tomato plants. These are now being field tested for resistance to Xcv. The environmental benefits of these plants could be substantial, as bacterial spot disease is presently controlled by spraying fields with massive amounts of copper and other toxic chemicals.

The multiple functions of the pgip gene family

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Although polygalacturonase-inhibiting proteins (PGIPs) were named for their ability to defend plant cells against fungal endopolygalacturonases (PGs), it turns out that they wear other hats as well. This is to be expected, Dr. De Lorenzo said, because PGIPs are made by genes in the leucine-

rich repeat (LRR) family, some of which play key roles in development while others confer resistance to pathogens. In addition to PGs, PGIPs interact with macromolecules including methylated pectins and membrane-associated lipoxygenases.

Dr. De Lorenzo, in collaboration with Dr. Fred Ausubel of Harvard, has been exploring the physiologic significance of these interactions in *Phaseolus vulgaris* and *Arabidopsis*. They have found significant redundancy in *pgip* gene families, with several genes encoding the same or related products. Using knock-out and over-expression mutants, they have begun to identify differences in recognition specificity, regulation, and function for *pgip* genes. One of the most exciting findings is that a *pgip* transgene generates a more heavily methylated pectin than the type found in normal cell walls. This may be useful in helping toughen plants against pests. The researchers have also used *pgip* transgenes to grow *arabidopsis* plants that are bushier than usual.

Proteolysis and the biology of antigen presentation

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Dr. Ploegh made the transition from plants to animals, describing an ingenious strategy that human cytomegalovirus (CMV) uses to escape detection by the immune system. Killer T cells are usually alerted to the presence of a viral invader when they spot viral antigens, bound to the MHC Class I complex, on the surface of infected cells. This display prompts CD8+ T cells to destroy infected cells. CMV sabotages this by making two proteins, US2 and US11, that keep the MHC Class I complex from reaching the cell surface.

Experiments in Dr. Ploegh's lab recently showed how these two proteins disrupt antigen presentation. US2 and US11 appear to grab newly synthesized MHC Class I products by their tails, which protrude from the endoplasmic reticulum (ER), and drag them into the cytosol. Being ripped from the ER in an untimely fashion, these complexes are viewed by the cell as mis-folded proteins; ubiquitin marks them for destruction and they are whisked off to the proteasome and shredded. There are some other actors in this plot, such as unknown proteins that strip off ubiquitin immediately before proteolysis, and the researchers are still looking for them.

CMV's wiles may help explain why this virus can infect such a wide range of cell types, especially in patients with AIDS or other forms of immune suppression. In the long term, experiments such as these may contribute to the design of superior gene therapy vectors, which might be able to evade the immune system en route to their targets.

Session 5: Cell Cycle, Senescence, Programmed Cell Death

Overview

Nothing goes on forever, including the capacity of a normal, well-nourished cell to keep dividing. When a cell becomes senescent, it stops dividing but remains metabolically active for a time, so that it gradually fades away. Programmed cell death is a dramatically different scenario, in which healthy cells act decisively to commit suicide. When senescence and programmed cell death occur at the right time and place, they are an integral part of the life and death of a normal organism. When they occur inappropriately, however, developmental abnormalities or disease can result.

Senescence and programmed cell death are complex phenomena that require a veritable symphony of intracellular signals and processes. Some of the individual contributors were examined by presentations in this session. The first paper described how temperature-sensitive small molecules can be used to selectively block the transport of materials from one organelle to the next. The second concerned programmed death in plant cells, which turns out to bear a surprising resemblance to apoptosis in animal cells. The genetic control of premature senescence in acute promyelocytic leukemia was the focus of the third, and the session ended with an update on the dynamics of LDL-receptor binding.

Presentations

Chemical genetics of membrane traffic

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Dr. Kirchhausen's laboratory specializes in clathrin, a protein involved in the formation of vesicles that sort and transport materials from the membrane to intracellular organelles. Several years ago, he and his colleagues used X-ray crystallography to solve the structure of clathrin. But they've had less success using yeast knock-out models to analyze its interactions with certain proteins. Chemical genetics offered them a new way to identify substances that can selectively block clathrin's biosynthetic pathway.

Chemical genetics is a novel method for identifying small molecules that disrupt gene or protein function. A fluorescent microscope is used to detect activity in a sample tray with nearly 400 wells, each containing whole cells to which candidate molecules and markers have been added. Dr. Kirchhausen used a temperature-sensitive glycoprotein whose migration from ER to golgi, then from golgi to membrane, can be controlled by temperature manipulation. This enabled the researchers to visualize the activity of different chemicals at selected points in biosynthesis.

Dr. Kirchhausen's team screened approximately 10,000 chemicals, and identified more than two dozen that disrupt the clathrin pathway. They found 2 chemicals that block the exit of newly-synthesized proteins from the ER, and 6 more that block exits from the golgi to the membrane. Eight others altered the structure of the golgi in various ways. The investigators were surprised to find 10 agents that could induce the formation of vacuoles in human cells, and to see that this process could be reversed without damaging the cells.

PML regulates p53 acetylation and premature senescence induced by oncogenic ras

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When the p53 gene functions normally as a tumor suppressor, it induces cellular senescence in response to oncogenic signals. Although the activity of the P53 protein is modulated by protein stability and post-translational modification, including phosphorylation and acetylation, exactly how the p53 gene is activated by oncogenes in the first place has been a mystery.

Now Dr. Pearson and his colleagues report that a tumor suppressor gene called *PML*, first identified in a mouse model for acute promyelocytic leukemia, acts in concert with *p53* to induce senescence. This gene appears to regulate *p53*'s response to oncogenic signals from Ras. Expression of this oncogene causes *p53* to accumulate and *PML* expression to increase, *PML* over-expression acetylates *p53* at lysine-382, and this makes *p53* biologically active. The outcome is senescence.

The researchers have also shown that Ras stimulation causes *p53* and the acetyltransferase CBP to form a trimeric *p53*-*PML*-CBP complex within the nuclear bodies, a site where *PML* occurs even in normal cells. Further evidence for *PML*'s role comes from knock-out experiments, which showed that *PML*-/- fibroblasts lose Ras-induced *p53* acetylation, *p53*-CBP complex stability, and senescence. These data establish a link between *PML* and *p53* and indicate that unless *PML* is on hand, signals from Ras will go unheard by the cell.

A system to explore programmed cell death in plant-pathogen interactions

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The hypersensitive response is a classic defense strategy of plants, in which resistance genes trigger programmed cell death (PCD) at the site of a pathogen invasion. In order to study signal transduction in this form of cell suicide, Dr. Stone and her colleagues developed a pathogen-free system they could use to trigger focal cell death in *Arabidopsis*. When they treated *Arabidopsis* with fumonisin B1 (FB1), a fungal toxin, the resulting lesions had the hallmarks of the hypersensitive response, including accumulation of phenolics, callose, and camalexin, production of reactive oxygen intermediates, and induction of pathogenesis-related gene expression. Although this model yielded cleaner data than they could have gotten using a whole pathogen, Dr. Stone's team thought they could do better still if they used an even simpler model.

They switched to using protoplasts, which are plant cells stripped of their walls, grown in culture. When these cells were challenged with FB1, the resulting cell death was consistent with PCD in *Arabidopsis*: it was dependent on *de novo* transcription, translation, and protein phosphorylation. Dr. Stone also observed that salicylate-, jasmonate- and ethylene-dependent pathways contribute to FB1-induced PCD, as indicated by FB1 susceptibility of mutants.

To identify other factors contributing to FB1-induced PCD, they selected FB1-resistant (*fbr*) mutants by sowing seeds on FB1-containing agar media. In this hostile environment, two resistant mutants, *fbr1* and *fbr2*, were able to grow. When these mutants were challenged with a different bacterial pathogen, a type of *Pseudomonas syringae* pv. *Maculicola* that expresses the avirulence gene *avrRpt2*, they exhibited no resistance. However, *fbr1* and *fbr2* displayed enhanced resistance to an isogenic strain that did not express *avrRpt2*. These results indicate that this protoplast system can be used to reveal mutants with pathogen phenotypes, and suggest that triggering host PCD is a common feature of compatible plant-pathogen interactions.

Structure of modular elements of cell-surface receptors

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The LDL receptor (LDLR) is the primary mechanism that animal cells use to take up particles of low-density lipoproteins from the blood. Healthy cells make LDL receptors and insert them into the plasma membrane when they need cholesterol for membrane synthesis. This normal process is disrupted in people with familial hypercholesterolemia (FH), who inherit defective genes for the LDLR. Because these mutated receptors cannot bind circulating cholesterol, abnormally high levels of lipoproteins accumulate in the blood and these individuals are at high risk for coronary artery disease.

Many researchers have studied the amino-terminal domain of the receptor, which is responsible for binding LDL and consists of seven tandemly repeated LDL-A modules. Each LDL-A module is ~40 residues long, and contains six cysteine residues engaged in three disulfide bonds. The fifth of these modules is regarded as being most critical for LDL binding, but the specific contact points between LDL and its receptor were unknown.

In his opening remarks to more than 100 researchers who gathered for the Symposium, HMS Dean Joseph Martin noted that it has been only three years since the Foundation began stimulating collaboration among Harvard's six basic science departments and between scientific centers in Italy and HMS. In his view, the Foundation has succeeded on both fronts. Today, its generosity supports not only the annual Symposium but also a broad range of joint ventures, including exchanges of technology and personnel and an increasing number of small seminars. Armenise-sponsored research is conducted at five Italian institutions: the European Institute of Oncology in Milano, the University of Padova, the Institute for Cancer Research and Treatment at the University of Torino School of Medicine, the Dipartimento di Ricerca Biologica e Tecnologica (DIBIT) at Scientific Institute San Raffaele in Milano, and Universita' Di Roma La Sapienza. At Harvard, the four Armenise centers are structural biology, neurobiology, cell signal transduction, and human cancer viruses. Representatives from all the Armenise programs participated in the 3rd Annual Symposium.

Impressed with the productivity of the Italian centers, the Foundation recently invited all of them to apply for a two-year extension of their original award. Dean Martin announced, during his opening remarks, that on the previous day the Foundation's board voted to extend funding for all five. ""This is very gratifying for the Italian part of the enterprise,"" Dr. Jacopo Meldolesi of DIBIT said in response to Dean Martin's news. The Armenise funding brings not only money, he said, but also scientific challenge and the opportunity to develop more mature, ongoing relationships with collaborators.

A considerable exchange of human talent is already underway. For example, Dean Martin cited the recent move of Armenise fellow Andrea Musacchio from HMS to the European Institute of Oncology, where he will lead a new structural biology department. In Dr. Daniela Pietrobon's closing remarks, the Padova researcher reminisced about three years she spent at HMS early in her career and credited the Foundation with making it possible for two young scientists from her group to follow in her footsteps. The Armenise grant has been invaluable for the support of young investigators and the strengthening of laboratory infrastructure, she said, and other scientists agreed.

The strength of the 3rd Annual Symposium's scientific program is one measure of the Foundation's success in fulfilling its mission. The event drew 112 basic scientists, slightly more than two-thirds of them from Italy. This year there were 46 poster presentations and 20 formal lectures grouped into five sessions:

Symposium Topics

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- Neurobiology I

- Biomedical Research

- Plant Defense/Pathogenesis

- Neurobiology II

- Proteolysis/Apoptosis/Cell Cycle

-

Session 1: Neurobiology I

Overview

So many Armenise-sponsored investigators are engaged in exciting neurobiology research that this year's Symposium devoted two full sessions to this vital and fast-moving field. The conference's opening session introduced four issues that are fundamental for understanding neuronal and brain function, according to Dr. Jacopo Meldolesi, who moderated the program. As distinct as these areas of research are, all are in fast-moving areas where even more interesting news can be expected in the near future.

The first presentation explores how the human nervous system transduces smell from the binding of odorants to special receptors in the nose to the transmission of information to the brain. The second speaker homed in on a large family of receptors, the SEX/plexins, that in mammals appear to play essential roles in the development of nerve tissue. The focus shifted to the cell membrane for the third talk, which introduced a new process of membrane regulation in neurons that molds the excitability of those cells. The final paper dealt with a fascinating transcription factor that appears to exert itself during embryogenesis by suppressing neuronal fate determination.

Presentations

Deconstructing smell

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Smells have shapes. But instead of having one receptor dedicated to smelling bananas and another to pine trees, Dr. Buck's research demonstrates that individual odorant receptors (ORs) recognize specific parts of an odorant's structure, rather than the whole molecule. The olfactory system combines data from numerous ORs to determine what it's smelling, which explains how humans and other mammals can discriminate an enormous variety of odors as well as pheromones (chemicals that elicit sexual or other innate behaviors). This combinatorial coding scheme relies on about 1,000 ORs expressed by the millions of sensory neurons in the olfactory epithelium lining the nose. Each neuron expresses one OR gene and neurons expressing the same OR are scattered throughout one of four nasal zones, an arrangement that decentralizes information processing. When a volatile chemical enters the nose, different parts of its structure are recognized by scattered neurons equipped with various ORs. Axons from these neurons travel to the olfactory bulb, where input from neurons with the same OR-no matter where they may be located in the nasal epithelium-is channeled into specific glomeruli with fixed locations, much as scattered light is focused by a lens.

The vomeronasal organ, where the subconscious detection of pheromones begins, is equipped with two distinct families of candidate pheromone receptors, one of which was first identified by Dr. Buck's laboratory. Inputs from these receptors appear to be processed separately from ORs, ensuring that signals for primal behaviors such as mating and aggression won't be confused with commonplace smells.

In recent experiments, Dr. Buck and her colleagues have combined calcium imaging and single cell RT-PCR (a technique for amplifying parts of the genome that encode proteins) to identify ORs for odorants that people perceive quite differently even though they are structurally similar. The combination of receptors that codes for a floral aroma, they found, may overlap substantially with the combination that picks up a rancid smell-but the two will be channeled to different glomeruli in the olfactory bulb. These observations of single cells support three important conclusions: a single OR recognizes multiple odorants, a single odorant is recognized by different ORs, and most odorants can be detected by different combinations of ORs.

The human Sex/plexin gene family encodes surface receptors for semaphorins and controls cell repelling cues

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Several years ago, when Dr. Tamagnone discovered an unusual family of receptor tyrosine kinase genes, he named the prototype SEX because it was located on the X chromosome. Additional research has expanded the universe of human SEX/plexins to include nine members grouped in four families. These genes were first identified in the nervous system, and they contain highly conserved sequences that are the same in nematodes as in humans. The SEX/plexins encode cell surface proteins homologous to the extracellular domain of the receptor tyrosine kinase that binds hepatocyte growth factor (HGF), a key growth factor in embryonic development. Early on, researchers determined that SEX/plexins appear to influence the migration of axon processes toward synapse formation. Now other roles are coming to light.

Dr. Tamagnone's team has observed that the extracellular domains of SEX/plexins (the parts that protrude from the cell surface) bear a strong resemblance to semaphorins, a large family of soluble and membrane-bound ligands. In Cell, they described an interaction between plexin-A and semaphorin-1 in Drosophila; since that report they have identified two semaphorins that bind to human SEX/plexin receptors. This is the first time that ligands for the SEX/plexins have been identified.

More recently, Dr. Tamagnone's laboratory has explored the possibility that the plexins are important in cell-to-cell signaling in epithelial and endothelial cells, in addition to their established role in neuronal development. When they used semaphorin protein as a probe, the researchers found that it bound to plexins that interact with neuropilins in cell signaling. Future experiments will explore whether semaphorin-binding plexins may also collaborate with neuropilins in signal transduction, acting via some novel cytoplasmic (inside the cell) structure.

Active chloride currents mold the electrical properties of the ""resting"" sympathetic neuron

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There's more to a resting neuron than meets the eye. Although the conventional view is that nothing much goes on when the membrane potential is below -60mV, Dr. Fesce and Oscar Sacchi, a biologist at the University of Ferrara, have detected a series of active conductances in the -60 to -80mV range. They now say that interactions between fluctuating currents of potassium and chloride mold the excitability of the cell and help determine how much synaptic input is required to trigger an action potential.

Working with the isolated, intact superior cervical ganglion of rats, Dr. Sacchi used a two-electrode, voltage-clamp system to characterize the conductances and electrical properties of these cells. Dr. Fesce used these experimental measurements to build a computational model of a complete sympathetic neuron, which revealed surprising interactions between a potassium current that acts just above the -60mV threshold and a subthreshold chloride current. A potassium current called I_A is activated at about -60mV and becomes fully activated at about -30/-40mV, causing transient depolarizations that rapidly relax to the initial value. During the upstroke of excitatory potential, Dr. Fesce said that this potassium current apparently gives rise to a counter current of chloride ions, which push toward repolarization. All this happens in a neuron that appears to be ""resting,"" with the result that a higher level of synaptic input will be required to trigger an action potential.

When a neuron is at rest, there is supposedly no net flow of ions across the plasma membrane. Nevertheless, Dr. Fesce found remarkable chloride currents that operate below -60mV. These currents respond to transient changes in membrane potential and return to a steady state after hundreds of seconds. Chloride conductance increases with depolarization, and at -60/-80mV the chloride equilibrium potential sustains a small inward current. Whatever happens to membrane

resting potential sparks changes in chloride redistribution and conductances, Dr. Fesce said, which challenges the notion that membrane potential reflects a simple balance between sodium and potassium.

Transcription factors in cell fate determination and differentiation

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Scientists have known for years that regulation of tissue-specific genes during embryonic development is controlled primarily at the level of transcription. Dr. Shi's experiments demonstrate that in Caenorhabditis elegans, certain enzymes are involved both in normal transcription and in abnormal cell growth. Nature has many strategies for releasing DNA from the compact chromatin packaging that enables it to fit into the cell nucleus. One such method is the attachment of an acetyl group to histones, which bind the chromatin package. Acetylation is determined by give-and-take between histone acetyltransferases (HATs), such as mammalian CBP and p300, and histone deacetylases (HDAs). Earlier work revealed that the adenovirus oncoprotein E1A cannot induce immature cells to divide endlessly so long as CBP/p300 are functioning, suggesting that these proteins are needed for normal cell growth and differentiation. More recently, Dr. Shi expanded on this idea by manipulating *cbp-1*, the gene encoding CBP-1, the C. elegans protein that corresponds to CBP/p300.

C. elegans may be a simple organism with less than 1,000 cells, ""but it does all the things that we do, pretty much,"" Dr. Shi said of his favorite model. Many of its 20,000 protein-coding genes are homologous with human genes. As in humans, cell fate is determined in part by transcription factors. When the researchers used a new technique called RNA-mediated Interference (RNAi) to inhibit expression of CBP-1, they saw undifferentiated embryos with no sign of normal morphology. At the stage when gut and muscle tissue are apparent, no such

differentiation had occurred. As expected, *C. elegans* genes that correspond to mammalian histone deacetylase appear to repress somatic differentiation. In a kind of tug of war, CPB-1 appears to promote endoderm differentiation by antagonizing the repressive effects of HDA. These experiments are the first to show how a homolog of the human proteins CPB and p300 functions in a live animal. These results also provide critical *in vivo* evidence that the histone acetylase activity of CBP-1 may be important for its biological activity. In addition to confirming Dr. Shi's hypothesis, these experiments yielded an unexpected result as well. When monoclonal antibodies were used to study the seemingly undifferentiated cells in worms without CPB-1 activity, those cells appeared to be neurons suggesting that neuronal differentiation may be a kind of default setting for *C. elegans* cells in the absence of CBP-1. Similar observations have been made recently in *Xenopus laevis*, suggesting that HATs may be a highly conserved, essential player in the differentiation of non-neural tissue.

Session 2: Biomedical Research

Overview

As chairman of the Foundation's Scientific Advisory Committee, one of Dr. Peter Howley's responsibilities is to shape the program for the annual symposium. As he organized individual abstracts to create this year's sessions, Dr. Howley discovered that some of the cutting-edge investigations sponsored by the Foundation aren't easy to categorize. He gathered four of them together for this session, which he dubbed Biomedical Research. The opening presentation dealt with viral vectors for gene therapy, followed by a tour of the enzymatic assembly lines that microbes use to grow antibiotics. The third speaker introduced the first mammalian mutation gene that appears to increase stress resistance and extend lifespan, and the fourth took his listeners inside a "chamber of doom" where proteins are destroyed. As disparate as these topics appear, Dr. Howley said that "they have a similar theme: all are in areas of research that interest me."

Presentations

New generations of lentiviral vectors for experimental and human gene transfer

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Penetrating cells is a do-or-die proposition for viruses, which can't replicate until they've gotten inside host cells. Researchers who are mindful of this special viral skill are seeking to use them as vectors for gene therapy or vaccines. Many viruses, however, aren't well suited to this task because they can only enter dividing cells, they don't penetrate very many cells, or they don't express the transfected gene at high enough levels. To get around these limitations, Dr. Naldini and his colleagues have designed hybrid lentiviral vectors capable of transferring and expressing genes in several rodent tissues *in vivo*, and in primitive human hematopoietic stem cells *ex vivo*. They have accomplished this by combining core elements of HIV-1, the pathogen that causes AIDS, with the envelope of a less harmful lentivirus called vesicular stomatitis virus (VSV). The safety profile for these vectors has improved as the researchers cut back on the amount of HIV genetic material they use, they've increased transgene expression through selective use of HIV *ltr* (long terminal repeat) and packaging signals, and VSV elements permit entry into a variety of cell types. Dr. Naldini's latest and safest vectors inactivate upon transduction and include only a minimal set of HIV genes.

In earlier experiments, his group demonstrated efficient delivery and sustained expression of marker genes by these vectors, both *in vitro* with human donor lymphocytes and *in vivo* when injected into the brains of adult rats. More recently, Dr. Naldini has been working with a mouse model for metachromatic leukodystrophy (MLD), an inherited liposomal storage disorder that in infants causes rapid, dramatic death as lipids accumulate in the central nervous system and other major organs. His team designed a hybrid lentivirus vector carrying the gene for ASA, the enzyme that is lacking in this condition, which they are testing in two ways. When the vector is injected directly into the brains of MLD mice, there is preliminary evidence that the transgene appears to express well and in a stable fashion. A second set of experiments involves *ex vivo* reconstitution of hematopoietic stem cells with the transgene, which the

researchers speculate will repopulate and replace the missing enzyme in animals. Early results appear promising, and additional research is underway.

Assembly line enzymology: the biosynthesis of polyketide and nonribosomal peptide antibiotics

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Because they live in a bug-eat-bug world, fungi and other microbes have evolved thousands of antibiotics that they use to disable their enemies and competitors. Because many of these substances disrupt only the functions of prokaryotic cells, leaving eukaryotic cells alone, doctors use them as invaluable weapons against microbes that cause human disease. All are made by a process that Dr. Walsh calls "assembly line enzymology." His laboratory seeks to understand the molecular logic of antibiotic assembly and "the flip side, by which bacteria produce nonribosomal peptide virulence factors, which allow them to infect you and me." Ultimately, the goal is to use combinatorial elaboration to assemble new antibiotics, block virulence factors, and fight growing problems of antibiotic resistance.

At a gestalt level, polyketide antibiotics such as erythromycin and tetracycline and peptides like penicillins and vancomycin look nothing alike. Yet all are templated natural products where assembly instructions come from the domain order of giant megasynthases. These molecules have "way stations," modules that enzymes, such as polyketide or nonribosomal peptide synthetases, use to initiate, elongate, and terminate the natural product chains. At some of these way stations, Dr. Walsh and his colleagues have been able to substitute one module for another, an accomplishment that could lead the way to combinatorial biosynthesis of new antibiotics. If investigators can build a parts list for the antibiotic assembly line, Dr. Walsh predicts that it will be

possible to swap modules around and create antibiotics that have not yet been made in nature, but which might be powerful weapons against resistant bacteria.

The p66^{shc} adaptor protein controls oxidative stress response and lifespan in mammals

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Although gene mutations that extend lifespan and enhance resistance to environmental stresses such as ultraviolet (UV) light or reactive oxygen species have been identified in *C.elegans* and other invertebrates, no such genes are known in mammals. In this presentation, Dr. Migliaccio announced that she and her colleagues have found a mutation in the mouse p66^{shc} gene that appears to have such properties. This gene is a splice variation of p52^{shc}/p46^{shc}, a cytoplasmic signal transducer involved in the transmission of mitogenic signals from tyrosine kinases to the Ras oncogene. Unlike p52^{shc}/p46^{shc}, which are known to cause malignant cell changes, p66^{shc} fortunately does not transform fibroblasts.

The researchers created a p66^{shc} knockout mouse that retained p52^{shc}/p46^{shc}, then *in vitro* subjected cells from that mouse to UV stress. After four days, wild-type cells were all dead, whereas the cells with the deletion were alive. Additional *in vivo* experiments showed the knockout mice to be more resistant to paraquat-induced oxidative stress than wild-type mice. The researchers suspect that p66^{shc} is part of a signal transduction pathway that regulates oxidative stress response, and that hypothesize that disrupting this pathway will protect against this well-known cause of aging. Further support comes from an observational study, in which a group of mice with a

double p66^{shc} deletion outlived those who were missing one copy and those with two normal copies of the gene.

Mechanisms of Protein Degradation within Eukaryotic Proteasomes

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For cells, an important part of viability is the prompt and appropriate degradation of intracellular proteins. In mammalian cells, large structures called proteasomes degrade proteins that have been marked for destruction by ubiquitin. Some of the fragments that emerge from this process are converted to amino acids; others are antigenic peptides that trigger cytolytic T-cell activity after presentation on MHC-class 1 molecules.

Unlike typical proteases, mammalian 20S and 26S proteasomes degrade proteins in a highly processive fashion that Dr. Goldberg describes as a "bite-chew" model. Once a protein substrate has been labeled by ubiquitin, an ATPase shepherds it into the proteasome's central "chamber of doom," where it will be unfolded and methodically chopped into small pieces. Each molecule is completely chopped up before the proteasome moves on to the next. Dr. Goldberg and his colleagues were surprised to find that all the products of this process are the same size, whether they started out as small polypeptides or big proteins, which they see as an indication that proteolysis continues until the products are small enough to diffuse out of the proteasome, then stops. About 99% of these fragments are smaller than 25 residues, with most in the 3-20 residue range. Only 10-15% of the products are 8-9 residues in length, the size required for MHC-class-1 presentation.

Eukaryotic 20S proteasomes contain active sites that cleave proteins in three distinct ways: two cut like chymotrypsin, two like trypsin, and two like caspase. The researchers were surprised to find that instead of acting independently, chymotrypsin appears to ""bite"" the substrate first, which initiates ""chewing"" by the other processing sites. Caspase substrates signal chymotrypsin when it's time to take the next bite. The resulting process is a highly efficient method for destroying abnormal proteins and for alerting the immune system to the presence of viruses and other undesirables. In the future, proteasome inhibitors may hold promise as treatments for cancer and other human diseases.

Session 3: Plant Defense/Pathogenesis

Overview

The more that scientists learn about how plants protect themselves against disease, the more parallels they see between plant and mammalian defense systems. Much of what is known about how plants respond to pathogen attack has come from studies of *Arabidopsis thaliana* or rice. This type of research will get a boost in 2000, when the genomes of these model systems are expected to be fully sequenced, Dr. Brian Staskawicz predicted in his introduction to this session.

Plants have many enemies, including bacteria, fungi, viruses, and nematodes. Agriculturists have been developing disease-resistant plants since the turn of the century, relying almost entirely on classical breeding, hybridization, and recurrent selection for desirable traits. Long before genes could be isolated, it was obvious that a single resistance gene could make the difference between a bountiful harvest and a failed crop, Dr. Staskawicz noted. After genetic mapping and map-based cloning became available, scientists began cloning specific genes for disease resistance. Many plant resistance genes have leucine-rich repeat(LRR) domains, which are important for recognizing non-self proteins in the environment and setting in motion a signal transduction pathway that ultimately leads to defensive action. The machinery for recognition and response closely resembles the mammalian immune system.

In recent years, studies of bacteria that prey on plants have revealed that they share many characteristics with human pathogens, including similarities in effector proteins known as

virulence factors. Now that both host plants and their pathogens can be genetically manipulated, scientists are going to find better ways to give plants an edge over their enemies. ""The field of plant pathogenesis and defense is exploding right now,"" Dr. Staskawicz said.

Presentations

Exploiting polygalacturonase-inhibiting proteins (PGIPs) to engineer novel plant receptors

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Plants can be resistant to disease only when there is a match between a plant resistance gene and an avirulence (Avr) gene in the pathogen. Resistance genes are thought to code for receptors that recognize specific Avr products. With one exception, all the plant resistance genes that have been identified have leucine-rich repeats (LRR), which encode specific receptors for a wide variety of Avr proteins. Dr. De Lorenzo suspects that recognition of specific pathogens hinges on a hypervariable region in resistance genes that codes for variations in a specific region within LRR proteins. This is theoretical at present, however, because her team has not yet observed a direct interaction between LRR and Avr proteins. Polygalacturonase-inhibiting proteins (PGIPs), present in the cell wall of many plants, belong to the large family of LRR proteins and are structurally similar to other known products of resistance genes. PGIP recognizes endopolygalacturonases (PG), enzymes that disease-causing fungi use to breach the cell walls of plants. PGIPs and PGs offer a unique opportunity to analyze how LRR proteins recognize specific attackers. Dr. De Lorenzo's laboratory

has been using site-directed mutagenesis to explore how the recognition capacity of PGIPs can be manipulated. In one experiment, alteration of a single amino acid residue caused a PGIP to lose function, she reported. Now her team is working with mutations that may increase recognition, seeking to create chimeric proteins that will enable plants to identify and resist a wider range of pathogens.

Plant extracellular matrix and development: targeting pectins

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Dr. Cervone's laboratory focuses on events in the plant cell wall, which is the organism's first line of defense against pathogenic invaders. For some years, they have pursued genetic mutations in the cell wall that may be important in both defense and normal development. When these proved elusive, the researchers decided to see if cell-wall mutations could be detected if they used well-characterized bacterial enzymes to manipulate extracellular matrix architecture. Dr. Cervone's team used Agrobacterium-mediated transformation to produce plants that overexpress polygalacturonase (PG) and polygalacturonase-inhibiting proteins (PGIPs). They were able to generate two kinds of transgenic plants: Arabidopsis and tobacco plants expressing PG from Aspergillus niger, and Arabidopsis, tobacco and tomato plants expressing PGIP from Phaseolus vulgaris. These alterations changed the pectins in the cells walls.

Although they expected that the morphogenesis of transformed plants would not be the same as normal ones, they were surprised when the altered plants grew much larger and more vigorously than the wild-type, a kind of plant version of super mouse. Another unexpected finding was that pectins from tomatoes with the pgip transgene exhibited a higher degree of methylation and acetylation than those isolated from non-transformed plants. This is consistent

with earlier findings that PGIP *in vitro* interacts with methylated pectins better than with non-methylated homogalacturonans, a preference that probably protects pectins from demethylation. Although much more research is needed, even at this early stage it is clear that plant transformation with PGs and PGIPs is a valuable tool for exploring how changes in pectin structure affect plant development, physiology, and defense.

Pseudomonas aeruginosa pathogenesis in diverse hosts

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Dr. Rahme's laboratory studies the molecular mechanisms underlying *Pseudomonas aeruginosa* pathogenesis in mammals. This is a medically important opportunistic bacterium that infects burn and trauma patients, as well as other immunocompromised individuals, and it is the leading cause of death in people with cystic fibrosis.

She and her collaborators have shown that a novel strain of *P. aeruginosa* uses the same subset of virulence factors to cause disease both in plants and in a wide range of animals, including humans. This important finding gave them the opportunity to use plants as a screening system for bacterial virulence factors, which once found could point the way to the identification of new antibiotics. This screening algorithm markedly decreases the use of laboratory animals, yet it generates data relevant to pathogenesis in mammals. In plants, Dr. Rahme's group has identified several novel *P. aeruginosa* virulence-related factors, the majority of them affecting the persistence and severity of infection. When these factors were tested in a mouse model that involves infection after a non-lethal burn, they made infections worse and accelerated sepsis development.

More recently, Dr. Rahme has been using *C. elegans* to identify avirulence mutations that enable the worm to feed on pathogenic *P. aeruginosa* and survive. She has found several mutations that appear to increase the host's ability to limit disease development, not just in worms but in mammals as well.

Molecular genetics of plant bacterial disease resistance

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Scientists have long anticipated a time when transgenic plants will be resistant to diseases that farmers now combat with chemical pesticides. Dr. Staskawicz and his coworkers will soon discover whether this moment has arrived for a type of bacterial spot disease caused by *Xanthomonas campestris* pv *vesicatorio* (XCV), which afflicts both peppers and tomatoes. A gene that produces durable resistance in peppers is being experimentally introduced into tomato plants, which have no natural resistance to this pathogen. It took Dr. Staskawicz' lab more than 8 years to nail down the function of this gene, isolate it, and prepare it for use as a transgene.

This work grew out of a broader inquiry into the molecular genetics of disease resistance, which showed that plants are protected against XCV-caused spot disease when the bacterium carries the avirulence gene (actually an effector protein) *avrBs2* and the host plant has the resistance gene *Bs2*. A series of genetic and biochemical experiments demonstrated that the *Bs2* gene product recognizes the business end of XCV—an effector protein that the pathogen needs to be at its most virulent. Having decided that *Bs2* was a good candidate for insertion into the tomato genome, Dr. Staskawicz and his colleagues set out to map and clone the gene—no trivial task in a genome four times as large as the human genetic endowment. The next step is to show protection in field trials, which are expected to begin this year.

Session 4: Neurobiology II

Overview

In this session, new scientific tools begin to pick apart some of neurobiology's venerable knots. For example, Dr. Elio Raviola said in his introduction, circadian biology used to mean watching how rats behaved when the light was switched on and off. The first paper described how molecular biology can pry open the mammalian circadian clock, so that some of its gears and springs can be spread out for inspection. The second presentation examined calcium pumps, certainly among the biggest and most important membrane transporters, and asked how they might be expressed during neuronal development. The next speaker tackled a classic problem, the ""inside-out"" migration of newly hatched neurons to their proper places in the cerebral cortex, and identified two major actors in this journey. Finally, the concluding speaker began to lift the veil on what could prove to be the master switch for neuronal exocytosis.

Presentations

Studies on the molecular mechanism of the vertebrate circadian clock

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Circadian clocks are endogenous oscillators that drive daily rhythms and physiology, and as such they probably represent an ancient and fundamental mechanism. In mammals, the central clock is located in the super chiasmatic nucleus of the brain, with independent clocks in each retina. Two years ago, investigators at Northwestern University identified the first mammalian circadian gene, which they called *Clock*. Further research showed that this gene encodes a presumptive transcription factor that is closely related to a family of proteins that mediate dimerization and bind DNA.

These findings immediately reminded Dr. Weitz of what his lab had learned about the workings of the *Drosophila* clock, where *per* genes make proteins that dimerize with the product of *tim* (timeless). This protein pair is transported to the nucleus where it somehow shuts down the *per* and *tim* genes until the proteins disappear and the genes turn on again. These findings motivated him to look for a similar feedback loop in mice. The search for a partner for CLOCK protein turned up BMAL1, which is co-expressed with CLOCK and PER1 at known circadian clock sites in brain and retina.

Additional experiments showed that CLOCK-BMAL1 heterodimers activate transcription from E-box elements, a type of transcription factor binding site, located adjacent to the mouse *per1* gene, and from an identical E-box known to be important for expression of *per* genes in *Drosophila*. If the CLOCK protein was mutated, however, it joined with BMAL1 to form heterodimers that bound DNA but failed to activate transcription. According to Dr. Weitz, CLOCK-BMAL1 heterodimers drive the positive component of *per* transcriptional oscillations, which appear to underlie circadian rhythmicity. This is the first time that binding has been proved to activate transcription.

More recently, transfection studies in mice have provided direct evidence that expression of the PER protein inhibits *Per1* gene activation by CLOCK-BMAL1. Protein interaction experiments and further analysis suggest that PER binding sequesters the CLOCK-BMAL1 heterodimer in a manner that keeps the transcription factor from binding to its E-box target site.

Calcium controls the transcription of its own transporters in developing cerebellar neurons

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Eucaryotic cells maintain a low intracellular concentration of free Ca^{2+} mainly by relying on a membrane-bound ATPase, PMCA, that serves as a high-affinity pump. Calcium is also exported from cells by a low-affinity $\text{Na}^+/\text{Ca}^{2+}$ exchanger (NCX). The PMCA protein has four isoforms that vary slightly in amino acid sequence, with PMCA4 being the best studied of these. In order to learn more about the others, Dr. Carafoli's laboratory made monoclonal antibodies to isoforms 1-3. Using these tools, they found PMCA2 and 3 only in brain tissue, whereas isoforms 1 and 4 appear ubiquitous.

In order to explore the role of PMCA isoforms 1-3 in neuron development, Dr. Carafoli and his colleagues used antibodies to track changes in these key carrier proteins in cultures of rat cerebellar granule cells (CGC). They found that PMCA isoforms 2 and 3, and a splicing variant of PMCA1 (designated as PMCA1CII) are strongly upregulated in the 3 to 5 days required for full maturation of the granule cells, whereas PMCA4 is much more rapidly downregulated. These effects occur at both the transcriptional and translational levels. Maturation of cerebellar granule cells requires the sustained influx of Ca^{2+} through L-type channels; when this was blocked with nifedipine the up-and-down regulation of PMCAs was abolished. At variance with the upregulation of PMCA 2, 3, and 1CII, the down-regulation of PMCA4 depends on increased levels of calcineurin, a relationship that was disrupted by immunosuppressive drugs. Dr. Carafoli also identified three isoforms of NCX at work in these cultured cells. NCX I and NCX III become slightly upregulated as the granule cells mature, whereas NCX II is strongly and rapidly down-regulated in a calcineurin-dependent way. The splicing variants of NCX1 also undergo a switch during maturation.

Expression of Ca²⁺ transporters may change because the cells need to gain better control over the increased Ca²⁺ influx required for the increased gene transcription that is integral to their maturation. Future research will explore the specific properties of the individual Ca²⁺ carrier proteins.

Function and dysfunction of cyclin-dependent kinase 5 in development and degeneration

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Neurons are born close to the inner surface of the neural tube and migrate outward to form the layers of the mammalian cerebral cortex. In these layers, neurons are grouped according to morphology. During cortical development, successive generations of neurons migrate in an "inside-out" fashion, with the first-born brain cells settling closest to home and later cohorts traveling just past them to form the next layer. Although this pattern is well-known, the factors that guide newborn neurons into place have been a mystery.

Over the past several years, Dr. Tsai has not only figured out how neurons migrate but also has uncovered connections between this phenomenon and Alzheimer's disease. Her experiments suggest that normal "inside-out" migration requires that cyclin-dependent kinase 5 work closely with p35, a regulatory protein. When she and her colleagues knocked out the p35 gene in mice, and when another lab independently knocked out the gene for cdk5, each produced mice with neurons layered "outside-in." Dr. Tsai's mice were defective but viable; the animals without cdk5 were not viable. She has since found evidence suggesting that the p35/cdk5 kinase complex facilitates "inside-out" migration by regulating actin cytoskeleton dynamics and reducing cell-cell adhesion, making it easier for freshly minted neurons to slip past the ones that have already settled in their appropriate layers.

When ischemia, hydrogen peroxide, or other means were used to stress the brains of mice, the animals converted p35 to p25, a protein that Dr. Tsai believes deregulates cdk5 and has no normal developmental function. Her team found massive accumulations of p25 and cdk5 in post-mortem brain samples from Alzheimer's disease patients, especially in the neurofibrillary tangles that are one major hallmark of the disease. Abnormally phosphorylated tau protein also abounds in these tangles. Future research focuses on the role of p25/cdk5 in apoptosis, and the possibility that this might be used as a target for treating neurodegenerative disorders.

Regulated secretion expression competence and multiplicity in neurosecretory cells

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Neurosecretion is the process by which cells express and release by exocytosis both the small vesicles, which contain classical neurotransmitters, and the large dense vesicles containing mixtures of amines, ATP, proteins and peptides. Because neurons and endocrine cells acquire secretory capacity during development and retain it, this is generally regarded as a stable trait that can be lost only in case of cell de-differentiation. Some years back, Dr. Meldolesi's laboratory developed what he characterizes as ""a neurosecretory cell that is incompetent for secretion."" Defective clones of pheochromocytoma PC12 cells appear phenotypically normal except that they lack the dense granules typical of neurosecretory cells; functionally they have lost the ability to secrete. These rat cells lack not only secretion products but also vesicle membrane proteins, including the vSNARE VAMP2, the plasmalemma tSNAREs, which are necessary for exocytosis, and various soluble regulatory proteins. The mechanism(s) sustaining the defect appear(s) to be at least in part post-transcriptional. When defective cells are fused with normal rat PC12 or with secretory human cells, or when they are transfected with one or more normal genes, neurosecretion returns

to normal levels. This implies the existence of genetic controls for exocytosis, and the nature of these mechanisms is now being investigated

Session 5: Proteolysis/Apoptosis/Cell Cycle

Overview

The topics covered in this session-protein degradation, cell cycle regulation, and apoptosis-may hold the keys to the development of better cancer therapeutics, Dr. Giulio Draetta said in his opening remarks. Unlike treatments for infectious diseases, which can be directed against features of the pathogen that aren't found in the human host, most cancer treatments must aim at molecules that are normally present. And this, of course, explains why so many anti-cancer treatments are so toxic to patients. In the future, molecular oncologists hope to have treatments that selectively kill cancer cells while leaving normal ones alone. Someday it may be possible for physicians to obtain a molecular fingerprint of the patient's cancer, then to select inhibitors that will moderate responses of the individual patient and the specific tumor. The presentations in session focus on signaling cascades that could ultimately prove relevant to the ultimate goal of finding less toxic treatments for cancer, Dr. Draetta said. He predicted that the next step will be looking for cross-talk among signaling pathways, in addition to exploring individual pathways, and that this will lead to even more ideas for high-specificity, low-toxicity treatments.

Presentations

The regulatory particle of the proteasome

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In his presentation on the first day of the symposium, Harvard's Dr. Alfred Goldberg described how cells use the ubiquitin-proteasome pathway to degrade proteins that would cause trouble if they were allowed to accumulate. Dr. Finley's work sheds more light on this crucial pathway. He used a yeast model to understand the opening and closing of the doorway through which ubiquitin-protein conjugates enter the lumen of the gigantic proteasome's 28-subunit core particle (CP). Once inside this chamber, tagged proteins are reduced to confetti. Although it would be easy to view the proteasome as a monolith, it is actually formed by the association of the CP with the 19S regulatory particle (RP), which sits over the CP channel and selects ubiquitin-conjugates for degradation. In addition, Dr. Finley's research indicates that the RP is a complex structure that fully unfolds substrates so they will fit through the 13Å-wide opening that leads to the CP lumen.

The yeast RP contains 17 subunits, 6 of them ATPases, and when viewed with an electron microscope this structure looks like a set of jaws that open and close to admit selected proteins. The lid is an 8-subunit subcomplex which can be dissociated from yeast proteasomes in vitro. The base is also an 8-subunit complex but it cannot be separated from the CP. The base contains all 6 proteasomal ATPases, which may both join the RP to the CP and propel some of the proteasome's targets into the CP for destruction, Dr. Finley said. By experimenting with various RP mutations, he and his colleagues have determined that rpt2, one of the ATPases found in the base, is needed to open the gated channel into the CP. The base is sufficient to activate the CP for degradation of peptides, perhaps indicating that it is competent to open the channel into the CP. However, the proteasome needs the lid to recognize ubiquitin-conjugates.

To determine whether the base of this assembly unfolds proteins in addition to opening the door, the researchers used citrate synthase (CS) as a model substrate. As they predicted, base ATPases acted as molecular chaperones for CS. Only after it was unfolded could CS be bound by the base, Dr. Finley said, and this reaction was independent of ubiquitin tagging. These data suggest that ubiquitin-protein conjugates are initially tethered to the proteasome via specific recognition of

their ubiquitin chains, followed by a nonspecific interaction between the base and the target protein, which is coupled to unfolding and translocation of the target protein into the CP.

Mitochondria in cell death: the (w)hole story

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Mitochondria are often referred to as the ""power plants"" of the cell because they specialize in synthesizing ATP. The most striking feature of these organelles is that they have a double membrane, which divides them into two compartments: the intermembrane space and the matrix space in the center of the structure. Mitochondria make few proteins in-house, and for the most part they import proteins from the cytosol which form complexes with mitochondria-made proteins. Since the early 1990s, mitochondria have been under close scrutiny as regulators of apoptosis, and as potential targets for therapeutic interventions directed at accidental or programmed cell death.

Some researchers suggest that the permeability transition pore (PTP) is a major player in mitochondrial apoptotic signalling. They postulate that when this high-conductance inner membrane channel expands to admit solutes, leading to tremendous swelling of the mitochondria, this may trigger release of intermembrane apoptosis-inducing factor and possibly of cytochrome c. In mechanistic terms, however, it is difficult to understand how this pore might be linked to the release of death factors by the organelle's inner membrane, Dr. Bernardi said. One barrier to understanding this process is that in vitro studies of cell-free mitochondria may not correspond well with in vivo events. Working with populations of mitochondria in suspension, Dr. Bernardi and his colleagues manipulated a variety of factors to try and mimic the in vivo opening and closing of the PTP. They found that depolarization always leads to opening of the pore, but that the reverse is not true. Further, they determined that a closed pore did not necessarily mean that the pump that drives ATP synthesis was also out of commission. More recently, the researchers devised a novel method for studying mitochondria in intact cells, which involves chemically blotting out background activity in the cell so that mitochondrial events stand

out. So far it appears that depolarization can result not only from opening of the PTP, but also from increased ATP demand or calcium influx. It also appears that GD3 ganglioside opens the pore and increases the likelihood of apoptosis, Dr. Bernardi said, and this is being studied further.

Structural communication in apoptotic pathways

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In order for a cell to die or commit suicide, its mitochondrial ""power plants"" must be shut down. But which of the many proteins involved in intracellular apoptosis pathways delivers the fatal blow to these organelles? The answer appears to be BID, a new protein that Dr. Wagner and his colleagues have identified in the complicated Fas signal transduction pathway. BID appears to link intracellular death signals to the mitochondria, where it sets in motion a chain of events that culminates with the activation of fatal caspase enzymes.

When the researchers used NMR spectroscopy to determine the structure of BID, they were surprised to find that this pro-apoptotic protein looks much like Bcl-xL, a protein known to inhibit apoptosis. Models of BID and Bcl-xL binding indicate that the two join easily in the presence of caspase-8, a death enzyme that loosens Bcl-xL's ordinarily tight structure. The complex of BID and Bcl-xL may interfere with the anti-apoptotic effects of APAF-1, which ordinarily binds with Bcl-xL. This sets the stage for lethal caspase activation that knocks out the mitochondria.

Regulation of cell cycle progression by the E2F transcription factors

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In mammalian cells, the retinoblastoma proteins (pRB) are key regulators of the cell cycle, serving as one of the main brakes on progress around the cell-division cycle. These proteins are essential for fundamental decisions about whether a cell should proliferate, differentiate, or undergo apoptosis. Of the numerous cellular proteins that interact with members of the pRB family, the best characterized are the E2F transcription factors. It is widely believed that the ability of the pRB family proteins to restrict cell proliferation depends on their ability to inhibit E2F transcriptional activity.

E2Fs are important for normal cell function, and dysregulation of these proteins has many consequences. Dr. Helin and his colleagues have generated cell lines expressing E2F-1, E2F-2, and E2F-3, each fused to the estrogen receptor ligand binding domain (ER), an innovation that makes it possible to manipulate ERE2F levels with hydroxy tamoxifen. Using this system, the researchers have found that activation of all three E2Fs can relieve the mitogen requirement for entry into S phase, and activation of the E2Fs leads to a shortening of the G0-G1 phase of the cell cycle by 6-7 hours. E2F can also induce apoptosis even in cells fed growth factors that would ordinarily sustain them, Dr. Helin reported. The researchers have also demonstrated that several genes containing E2F DNA binding sites are efficiently induced by the E2Fs in the absence of protein synthesis. More recently, Dr. Helin's laboratory has identified two novel targets for E2F transcription factors, both cell-division-cycle genes. One, *cdc25A*, is a tyrosine phosphatase essential for the activation of certain cyclin-dependent kinases and S-phase initiation; it is also overexpressed in many tumors. The second, *cdc6*, is not only needed for the cell cycle to advance but also is a very sensitive marker for cell proliferation; in some cancers it may be a marker for the aggressiveness of tumor cells. In the future, E2Fs can be used to scan the human genome for additional genes important for apoptosis, cell proliferation, or DNA replication, Dr. Helin said.

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